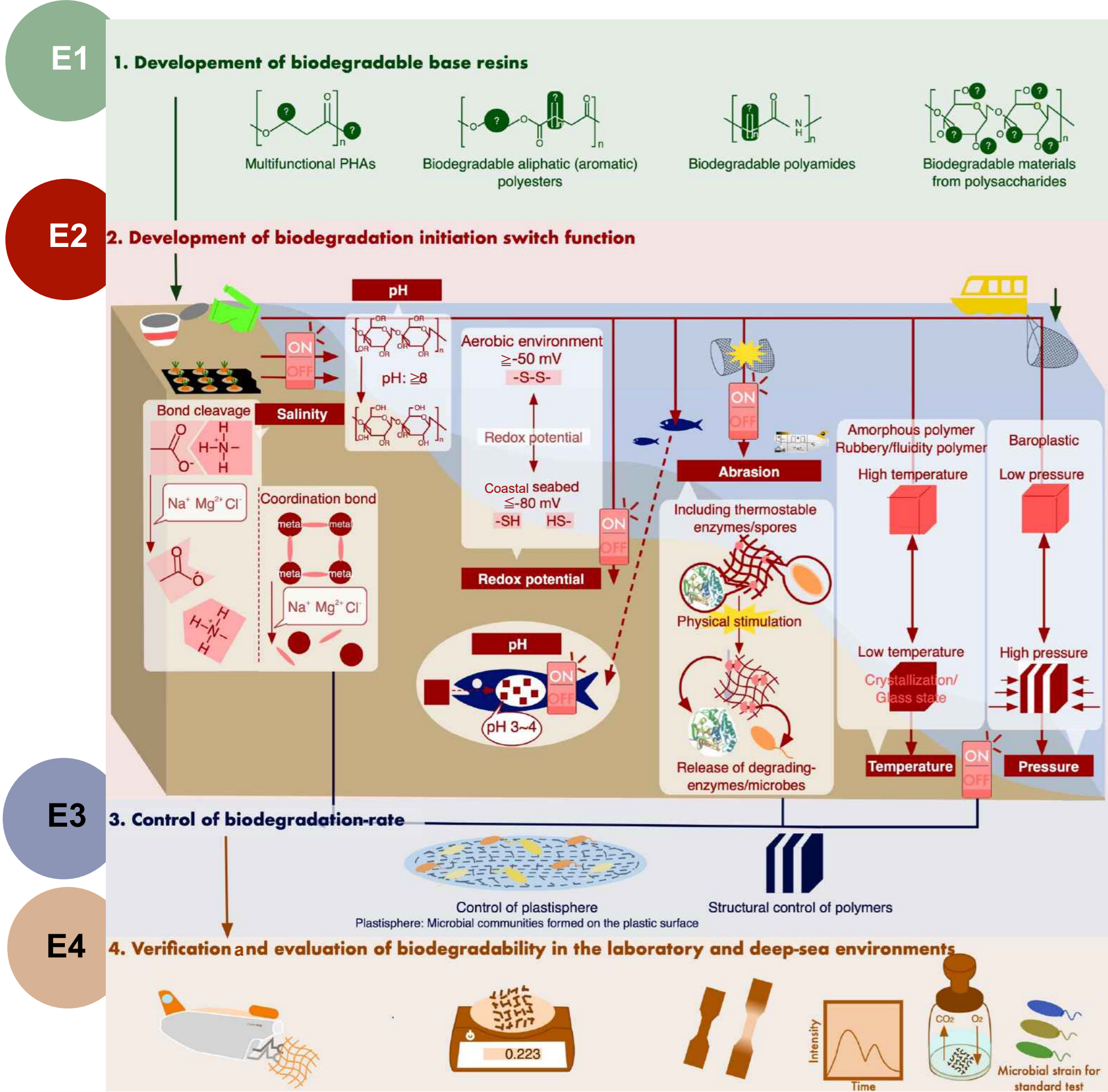
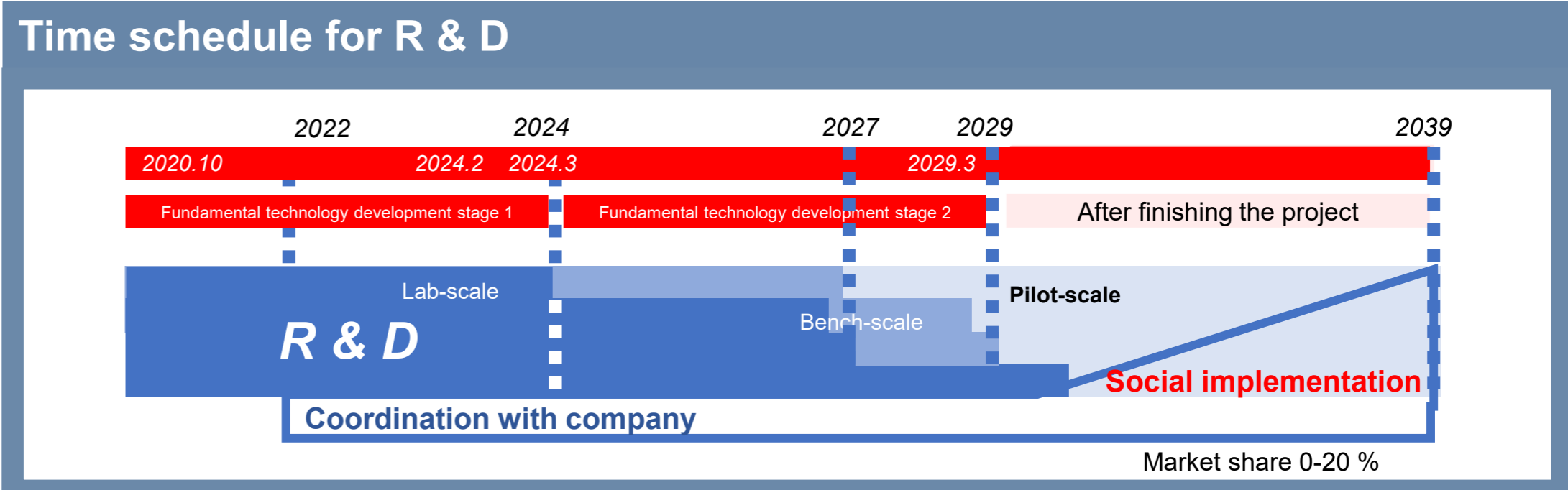


Overview and goal of our project

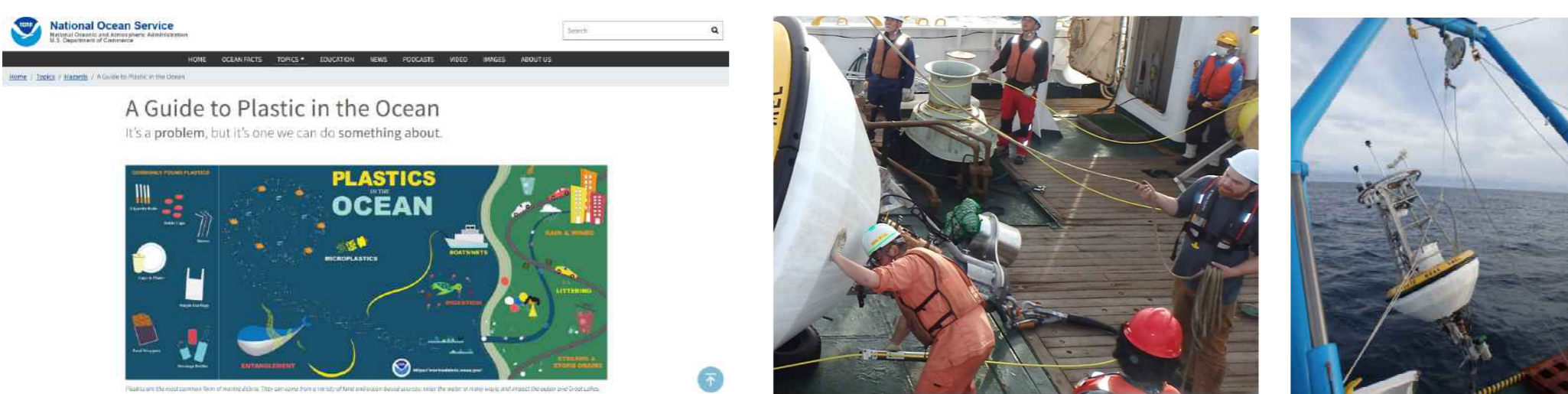


This project has the following goals for social implementation of development technology.

- ① We create three or more new marine biodegradable plastics that exhibit 90% biodegradability in seawater at 30 °C in six months after the switching function exerts.
- ② We demonstrate the biodegradability of these new marine biodegradable plastics having the switching function in marine environments, including deep sea.
- ③ We create new marine biodegradable base materials made from biomass and carbon dioxide.



International cooperation



We evaluated degradability of biodegradable plastics in the ocean surface layer in cooperation with the National Oceanic and Atmospheric Administration (NOAA). Buoys installed in the Great Pacific garbage patch is used as a test site for degradation of biodegradable plastics.

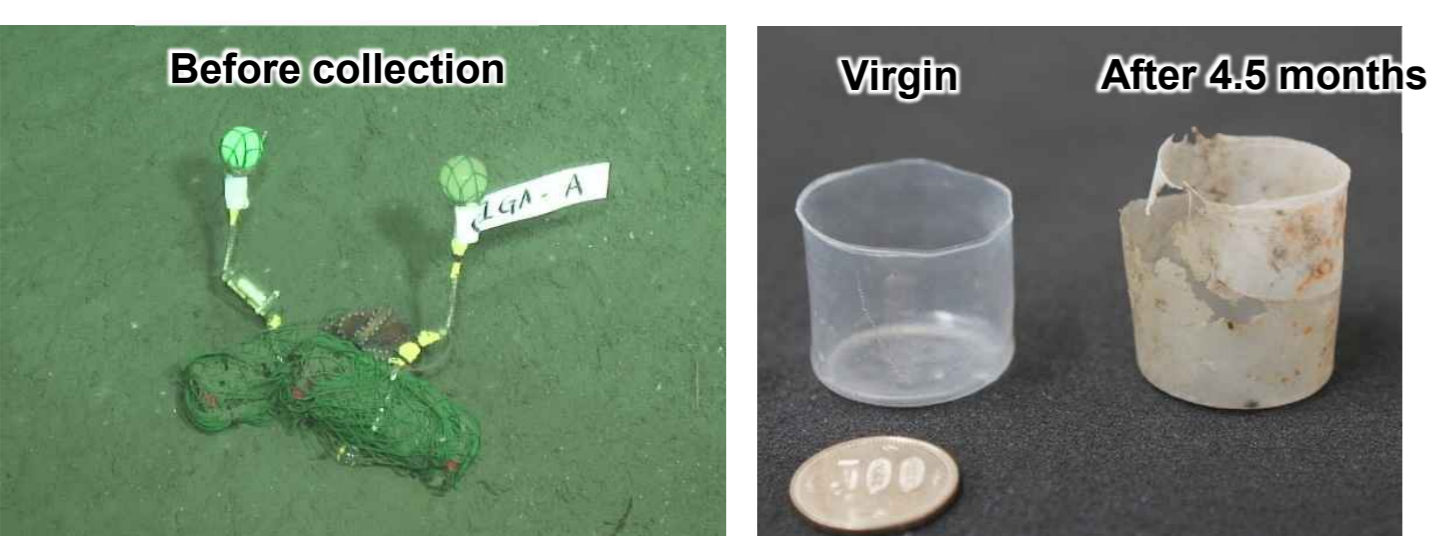
Science and technology dialogue with the public



As part of GIGA School x Deep Sea, a new biodegradable material was installed 855 m off Hatsushima Island with more than 24,000 elementary school students and the Minister of MEXT via a live online broadcast.

Media appearance

- TOYOBO Column: "Biodegradable Plastic R&D Leading to Solutions to Marine Debris Problems: Now and the future"
- Gunma Kankyo Keizai Forum 2022 20220225
- Biodegradable Plastic Standardization Consortium 20220119
- NTV Sukkiri 20220608
- NBC Radio program 20220622
- Gunma Future Innovation Conference 20220623
- Lecture on contemporary issues in Ibaraki prefecture 202107
- Shibukawa City Citizen's Environmental University 20201115



Above: The material before and after 4.5 months in deep-sea. Right: News site reporting the results of the material developed in this project and demonstrating on-site biodegradability.



Marine debris @Yokosuka

Fish net on beach @Sendai

Development of multifunctional microbial polyester: Base materials for installing switching functions

Glucose Carbon source + 4-Pentenoic acid → Recombinant *E. coli* → **P(3HB-co-3HPE)** (Unsaturated side chain) → **Installing switch function** → **Synthesize 200 g of 3H4PE polymer** → To install switch function

Sugars + 5-Hexenoic acid → Recombinant *R. eutropha* → **P(3HB-co-3HHx-co-3H5HE)** (Unsaturated side chain) → **Installing switch function**

Hexanoic acid → Intracellular saturation reaction → **P(3HP)-b-P(2HB)** (High toughness material)

End-modified PHA

Flask culture (2L) → Extraction

Other materials

Glucose → 2 Acetyl-CoA → PhaA → 4-Methylthio-2-oxobutanoic acid → LdhA → 2-Hydroxy-4-methylthiobutyric acid (2H4MTB) → HadA → 2H4MTB-CoA → PhaC → **New sulfur-containing PHA**

L-Methionine → Met derivative 2H4MTB → Oxidation (Sulfoxidation, Sulfonation) → Hydrophilization (79° to 60°) → Functionalization of thioesters

2HTMB ~30 mol%

Development of efficient synthesis method for new PHA from CO₂

CO₂ → Recombinant strain → **P(3HB-co-3HHx)[PHBH]**

The gas that is not used by the bacteria and is exhausted from the fermentation tank is returned to the gas storage tank.

Supply of raw material gas to the culture medium in the fermenter

PHBH productivity of various recombinant strains (flask culture)

Strain / Plasmid	Dry cell wt. (g/l)	PHBH Content (wt%)	3HB (mol%)	3HHx (mol%)
C.necator H16 (wild-type)	17.16	68.2	100.0	0
MF01/ pBPP-ccr _M J4a-emd	12.18±0.40	64.0±3.4	94.8±1.1	5.3±1.1
MF01ΔB1/ pBPP-ccr _M J4a-emd	10.65±1.35	61.7±4.6	52.3±6.2	47.7±6.2
MF01/ pBPP-ccr _M JAc-emd*	11.22±2.67	64.6±8.1	88.7±6.4	11.3±6.4
MF01ΔB1/ pBPP-ccr _M JAc-emd	8.52±1.00	67.8±1.8	87.1±2.3	11.1±1.3

* 3HHx 10mol% PHBH is known to have the best physical properties (~2020)

PHBH production test by jar culture of *C. necator* strain MF01/JAc

Dry cell (g/L)	PHBH (g/L)	3HB (mol%)	3HHx (mol%)	Culture time (h)	Productivity (g/L/h)
61.4	51.5	94.6	5.4	205	0.300
71.0	58.4	86.2	13.8	119	0.594

(Highest value as of November 2022)

Synthesis of polyesters from aromatic diols with resorcinol back-bone structure

Glucose → Fermentation and/or chemical process → **Resorcinol**

Catechin → Fermentation → **Resorcinol**

Resorcinol + Carboxylic acid → DIC, DPTS / DCM, 30 °C, 20 h → **resorcinol-based semiaromatic polyester**

2MR (2-methyl substitution), 5MR (5-methyl substitution), 25MR (2,5-dimethyl substitution)

GI: m = 3, Ad: m = 4, Pi: m = 5

resorcinol: R₁ = R₂ = H, 2MR: R₁ = methyl, R₂ = H, 5MR: R₁ = H, R₂ = methyl, 25MR: R₁ = R₂ = methyl

Succeeded in synthesis of polyesters from aromatic diols with resorcinol back-bone structure by using carbodiimide compounds. The products with high molecular weight (Mn>24kg/mol) could be obtained.

Marine biodegradable plastics produced from polysaccharides

Glucose → Fermentation and/or chemical process → **Paramylon**

Extraction Thermo-processing → Esterification, New processing procedure

Injection molding • Resistant to acids and alkalis • Better impact strength > PP

Melt-spun fibers • Processable without additives • High-strength

BOD biodegradation test

Using Seawater from Tokyo Bay

Successful development of new high-performance materials with controlled marine degradability from polysaccharides

Excellent biodegradability (2.1, 1.6, 2.3)

Non-biodegradability (2.6, 2.8)

Development of new cellulose-based transparent materials

Dissolving, coagulating, and drying cellulose gives transparent paperboard. PCT/JP2020/039874

Compositionally identical with paper but more functional.

A transparent cup made of cellulose

Gel → Cup → Drying → **Up to 2 mm thickness**, **Ductile**, **3D shaping**

A transparent straw made of chitin

Gel → Straw → Drying

The fragileness of chitin was overcome by the improved molding process.

Easy-coloring

Chitin straw: Complete decomposition in 2 months.

Cellulose cup: Complete decomposition in 10 months.

By the improved shaping process, the preparation of materials entirely made of pristine cellulose or chitin was successful.

Proof of concept of the transparent cup made of cellulose

Deployment of transparent cup made of cellulose at the deep-sea floor was broadcasted to the elementary schools in Japan in the presence of minister for MEXT.

Appeared in news23 (TBS) and 16 news papers.

Switching triggered by difference in salt conc.

Polymers introduced the ligands with dicarboxylic acid group

Polyvalent metals → **Introduction of metal ions** → **Formation of coordinate bonds** → **Addition of salts** → **Polymers with the terminal ligands**

Polymers with the terminal ligands → **Successed in introduction of ligands into the chain-ends of aliphatic polyesters**

Polymers with the terminal ligands + **Cl-Zn-Cl Zinc chloride** → **Linear polymers forming coordinate bonding between terminal ligands and metal ions**

Changes in molecular weights determined by GPC

Polymers with the terminal ligands/Metal ions after immersion in NaCl aq. for 2 days: $M_n=12,500$, $M_w/M_n=1.6$

Polymers with the terminal ligands/Metal ions before immersion in NaCl aq.: $M_n=51,100$, $M_w/M_n=1.7$

Polymers with the terminal ligands: $M_n=11,300$, $M_w/M_n=1.4$

Confirmed the cleavage of coordinate bonding of polymers by the immersion into NaCl aq. solution with a concentration of 2wt% or more.

Polymers with hydroxyl chain-ends (low MW)

Polyvalent metals → **Introduction of metal ions** → **Formation of cross-linkage** → **Addition of salts** → **Cleavage of cross-linkage via dissociation of coordination bonding**

CL → **PCL**

Cyclobutane tetracarboxylic acid dianhydride : CyBTC

Both-terminal-substitute molecules + **Chain-extended molecules (multiplicative molecules)**

Changes in molecular weights determined by GPC

PCL with cyclobutane tetracarboxylic acid + **Cl-Zn-Cl Zinc chloride** → **Cross-linking polymers forming coordinate bonding between ligands and metal ions**

PCL-CyBTC: $M_n=47,700$, $M_w/M_n=2.2$

PCL-CyBTC/Zn: $M_n=5,103,000$

Confirmed the partially cross-linking structure via formation of coordinate bonding between metal ions with tetrahedral coordination and the functional ligands of dicarboxylic acid group in polyesters

Switching triggered by wear (Endospore)

Endospore → **After wearing** → **Inflowing water and germinant substances** → **Vegetative cells** → **Biodegradation** → **CO₂+H₂O**

Degradation test of spore-containing PESu

● : Weight loss of spore-containing PESu film with Yeast extract (YE). ○ : Weight loss of PESu film with YE. △ : Weight loss of spore-containing PESu film without YE.

Switching triggered by difference in ORP

PBSDT → **Flexible film by melt-molding** → **Flowing out into marine environment** → **Biodegradation** → **Easily biodegradable compounds**

- PBSDT as an analogue of PBSA
- Reductive cleavage of disulfide bonding in low ORP condition
- Biodegradation of the reduced compound in marine

Reductive Cleavage → **Low ORP** → **HS-SH**

Switching triggered by wear (Enzyme)

Biodegradation triggered by wear and enhanced by embedded enzyme

Enzyme-embedded PLLA
Enzyme-embedded PBS
Enzyme-embedded PBSA
Enzyme-embedded PCL

Without enzyme: no degradation

Enzyme-embedded polyesters: High weight loss and degradation

Biodegradation in deep sea (Hatsushima, 15-20 °C, 3 and 9 month)

- Enzyme-embedded PLLA, PBS, PBSA, PCL were prepared
- Enzyme-embedded polyesters were successfully degraded in the ocean (Hatsushima, JAPAN)

Switching triggered by difference in pH

Biodegradation triggered by pH change and subsequent biodegradation of polysaccharide ester

Poly(hydroxy alkanate) (PHA) films coated by cellulose acetate (CA)

Released in the ocean → **Stable during use** → **Alkaline hydrolysis in the ocean with pH 7.5** → **Switch on** → **Biodegradation of PHA**

Deacetylation begins → **Biodegradation of CA layers** → **Rapid weight loss of PHA**

Holes on CA layer were observed. PHA layer started rapid degradation after CA degradation

- PHA films were coated with cellulose acetate (CA) layer.
- Biodegradation of CA layers was triggered by alkaline hydrolysis in the sea water and subsequent decrease in degree of substitution.
- Degradation did not occur in water with pH 7.

Biodegradation rate factors from materials science

High-strength and modulus PHA fiber

Microbial polyester fibers	Mechanical properties		
	Tensile strength /MPa	Young's modulus /GPa	Elongation at break /%
P(3HB)	1320	18.1	35
P(3HB-co-8 mol%-3HV)	1065	8.0	40
P(3HB-co-9 mol%-3HH)	552	3.8	48

Cannot be cut by pulling with all your might.

Biodegradation of fiber

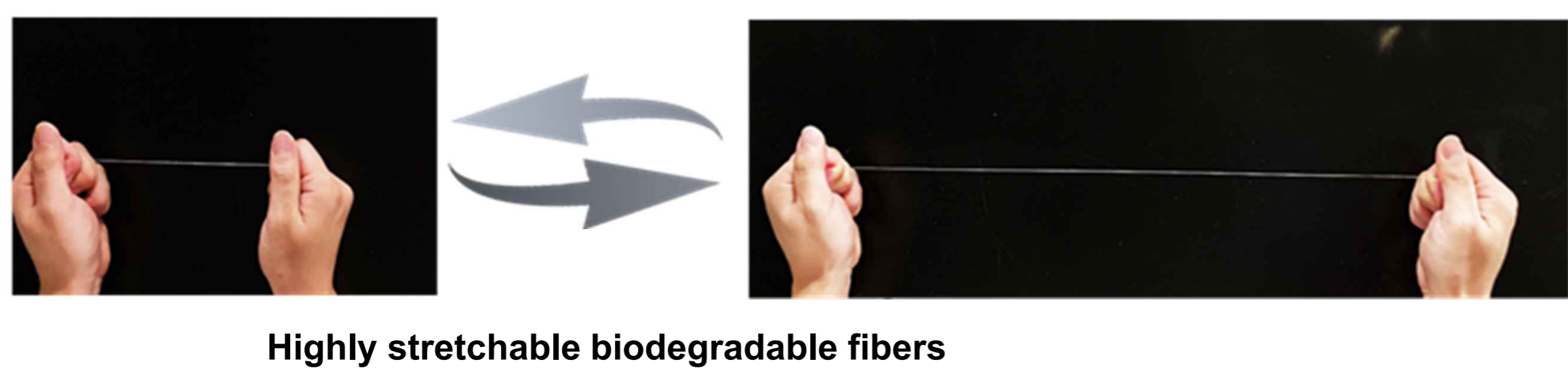
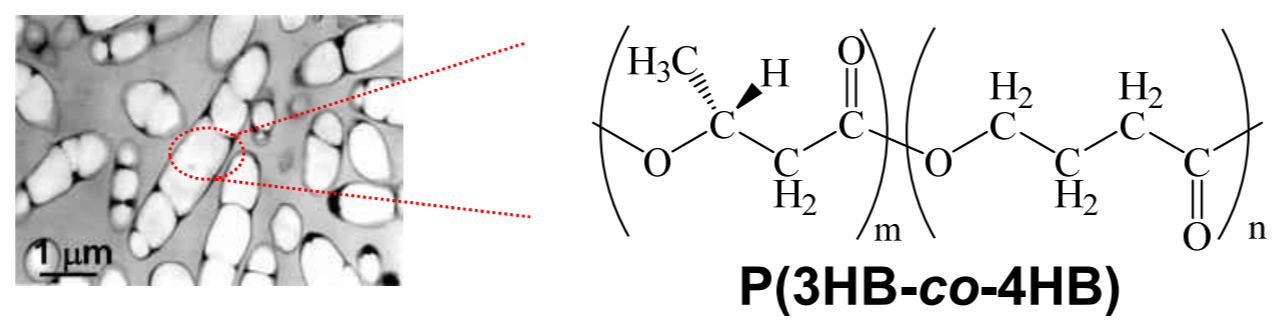
After 1 week
Surface
In biofilm
Cocci
Biofilm
Bacillus
Fiber

Bacillus on the fiber surface may degrade fiber using the enzyme.

- Degradation rate can be controlled by drawing ratio.
- Degradation rate is related to crystalline morphology.

Residual weight / wt. %
Time / days
Undrawn fiber
Drawn fiber
Rate difference

- Polyesters produced by microorganisms.
- P(3HB-co-4HB) in the one of the copolymers.



BOD biodegradability in seawater

BOD-biodegradability / %
Time / days
P(3HB-co-16 mol%-4HB) fibers
Cellulose (KC flock)
Formation of biofilm

1 Week
SEM image of the fiber during degradation
Microorganisms on fiber surface

Plastisphere: Microbial flora formed on plastic surface

Neuston in surface microlayer
On plastic
Marine snow
On macrophytes
On fish
On stone
Seawater

SEM image of plastisphere formed on the biodegradable plastic surface.

Microbial accumulation in the plastisphere
Coverage
GC content (%)
Deltaproteobacteria
Gammaproteobacteria
Bacteroidetes
Others
Alphaproteobacteria
Planctomycetes
Betaproteobacteria

Microbes with high abundance = Genome information of microbes involved in the biodegradation.

Elucidate the biodegradation mechanism of plastic and control the degradation rate.

Also investigate the plastisphere in non-oceanic environments.

Biodegradable plastics is exposed to pond.

Biodegradation rate control by controlling plastisphere

MDS1
MDS2
Microbial synthetic plastics
Biodegradable plastics in ocean
Chemosynthetic biodegradable plastics
Non-degradable plastics in the ocean

The plastisphere of Non-marine biodegradable plastics close to that of marine biodegradable plastics. → Improving biodegradability

Addition of 10% plastisphere control substance candidate to the biodegradable base polymer. The films were exposed to seawater and investigated weight loss and change in plastisphere.

nMDS2
nMDS1
Seawater
PHBV
PBSAのみ
13(PBSのみ)
13(PLAのみ)

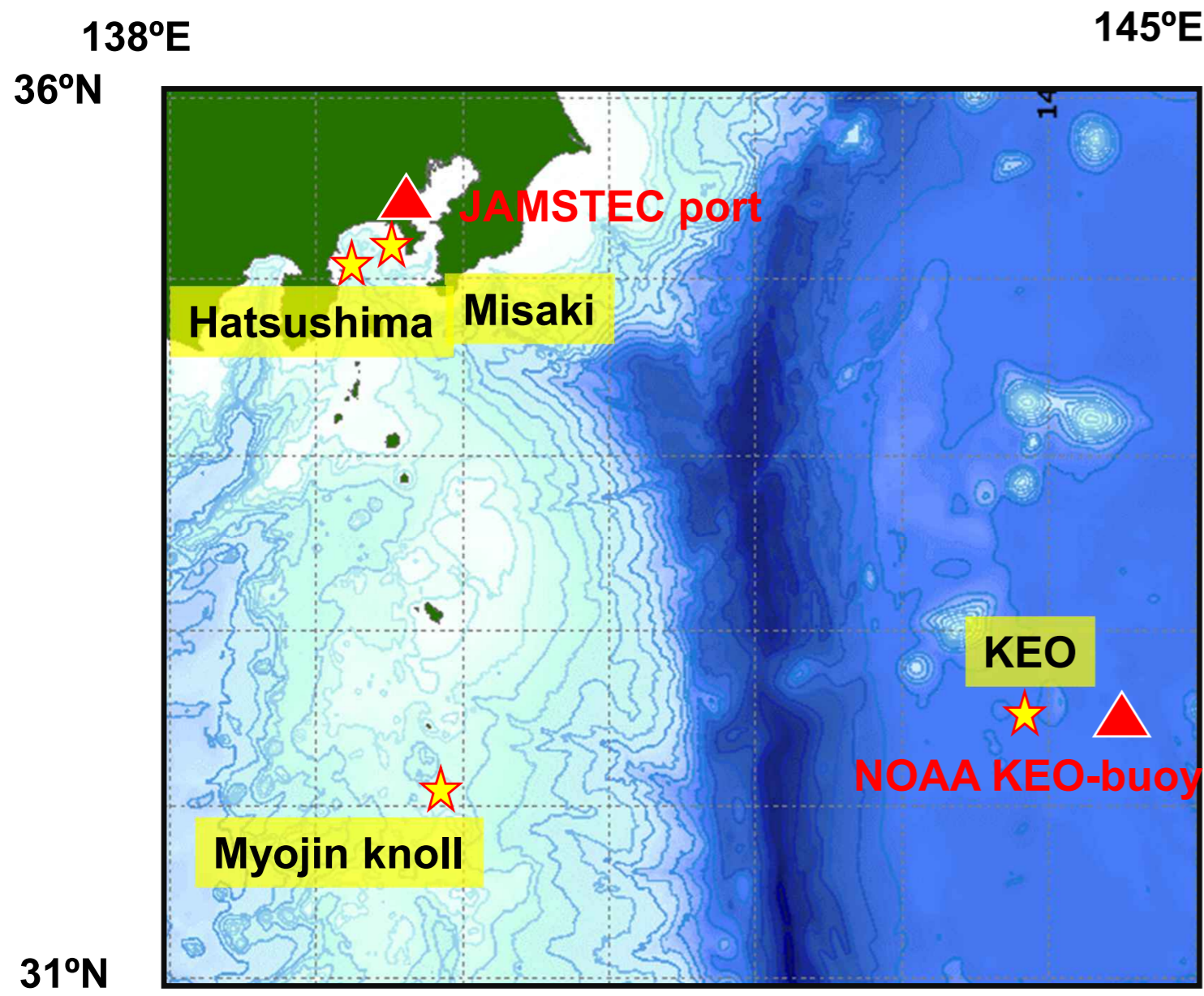
Plastisphere control substance candidates

Number	Substance
1	CE
2	A
3	CH
4	P
5	SH
6	HH
7	C
8	C
9	AN
10	AC
11	P
12	Y
13	Negative control

Effect on increase in the degradation rate.
PBSA No.2, No.5, No.6, No.7
PBS No.5, No.6
PLA No.2, No.7, No.9

Non-metric multidimensional scaling (nMDS) based on the Bray-Curtis index. Numbers in the plot indicate the type of substance. The area of the plot shows the biodegradation rate except for seawater.

In situ biodegradation tests of novel materials



- We carried out 6 cruises to test the biodegradability of newly developed materials on the deep-sea floor from 2020 to 2022. This is the only project that is testing biodegradability on the deep-sea floor in situ, where large amount of plastic debris are accumulating.
- We have also started biodegradability test at the surface of the North Pacific pelagic site.
- The recovered materials were examined with different chemical and physical tests, together with meta-omics approaches of the attached biofilms.



Deployment and recovery with the manned submersible



Plastic chambers deployed on the deep-sea floor

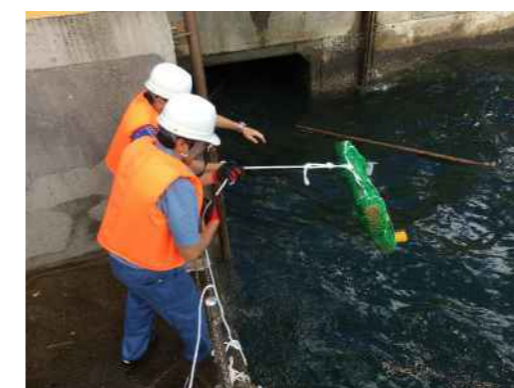


Plastic chambers deployed on the mooring buoy

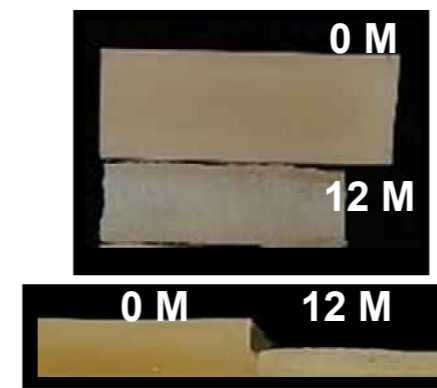
In situ biodegradation tests: in shallow water

Biodegradation of biodegradable plastics in shallow water

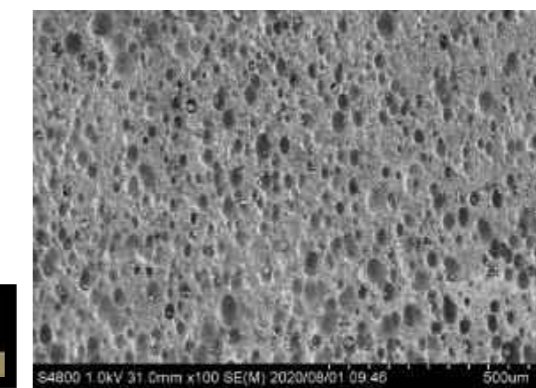
	Length (mm)	Wide(mm)	Thickness (mm)	Weight (g)
0 Month	30.0	10.0	4.0	1.30
12 Month	25.5	7.5	2.2	0.39
Reduction rate	15%	24%	45%	70%



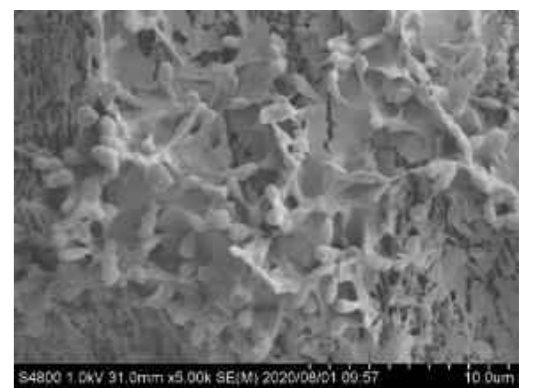
Setting



Size

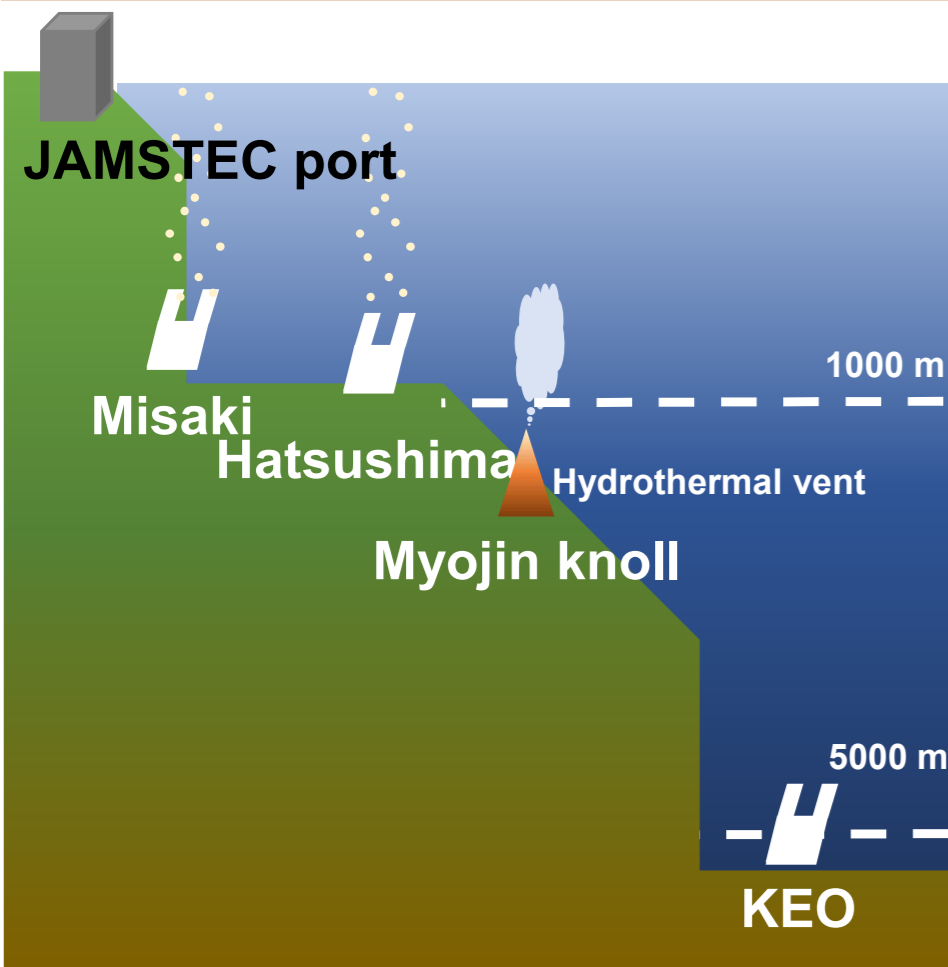


Surface



Microbes

In situ biodegradation tests in deep-sea



	Salinity	Temp.	DO
JAMSTEC port	28-29	23-26	
Hatsushima	34.5	3.6	1.24
Misaki	34.3	4.4	1.46
Myojin knoll	34.3	4.6	1.50
KEO	34.7	1.6	3.6-3.7



Biodegradable plastics



Samples after 4 months of installation



Recovery using a robotic arm



Chamber filled with biodegradable plastics



Recovery of samples and deep-sea water

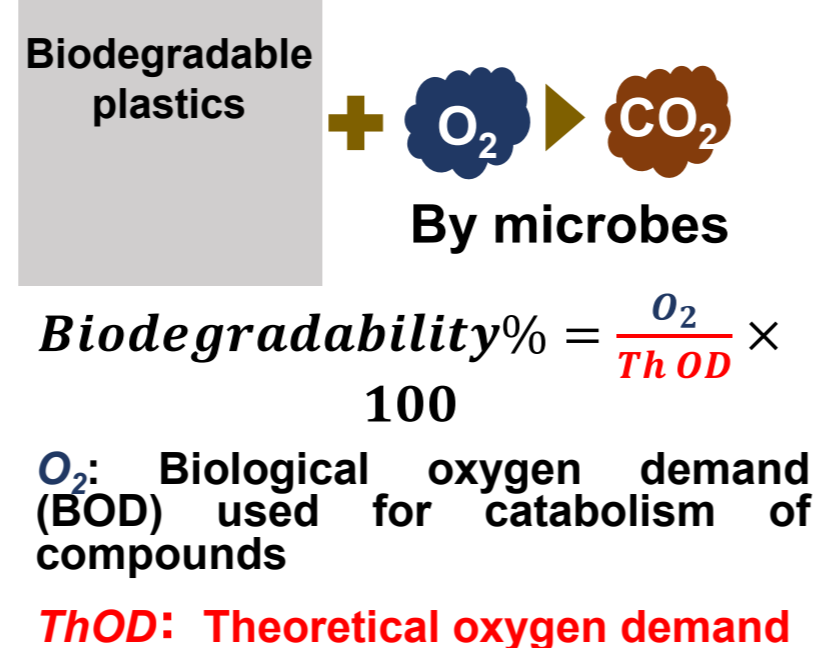


Collection of seabed soil

In vitro biodegradation tests of novel materials



BOD biodegradation testing



Optimization of in vitro test condition

① Biofilm development for obtaining plastic degraders

Laboratory (4 months)

PBSA and PBAT films were put in sea water of 4 sites.



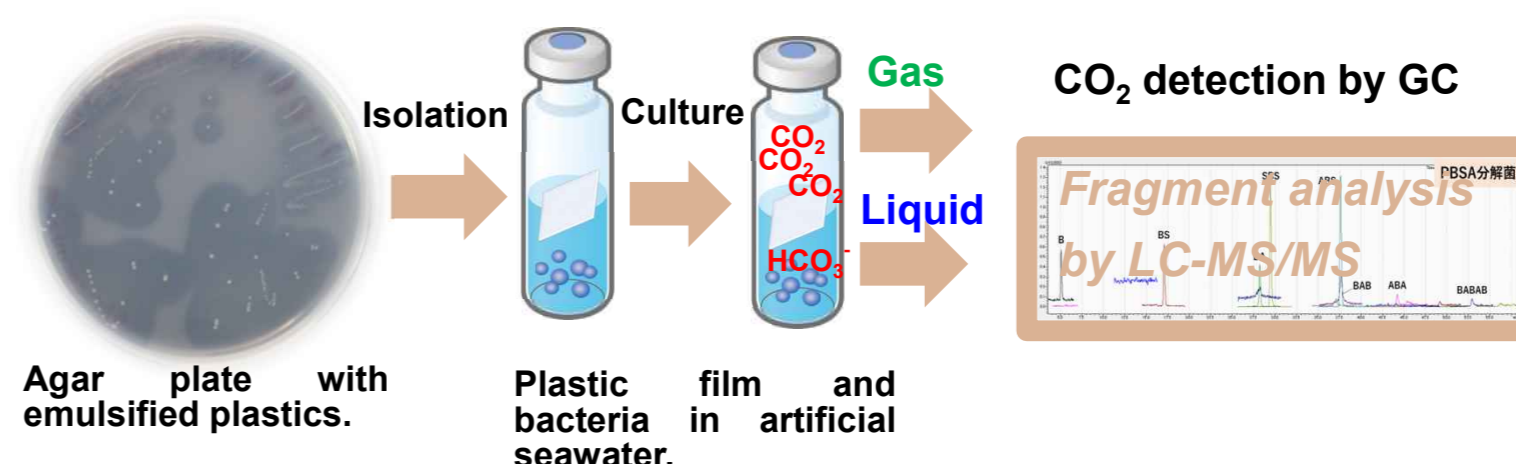
In the sea (5 months)

(Hiroshima, 10 m-depth)
Films of PBSA, PBS, PBAT, and PCL with film mounts were fixed in the fixture.



• PBSA/PBAT degraders (230 strains) were isolated from plastisphere.

② Degradation property analyses of degraders on each material and degrader selection for rapid test method

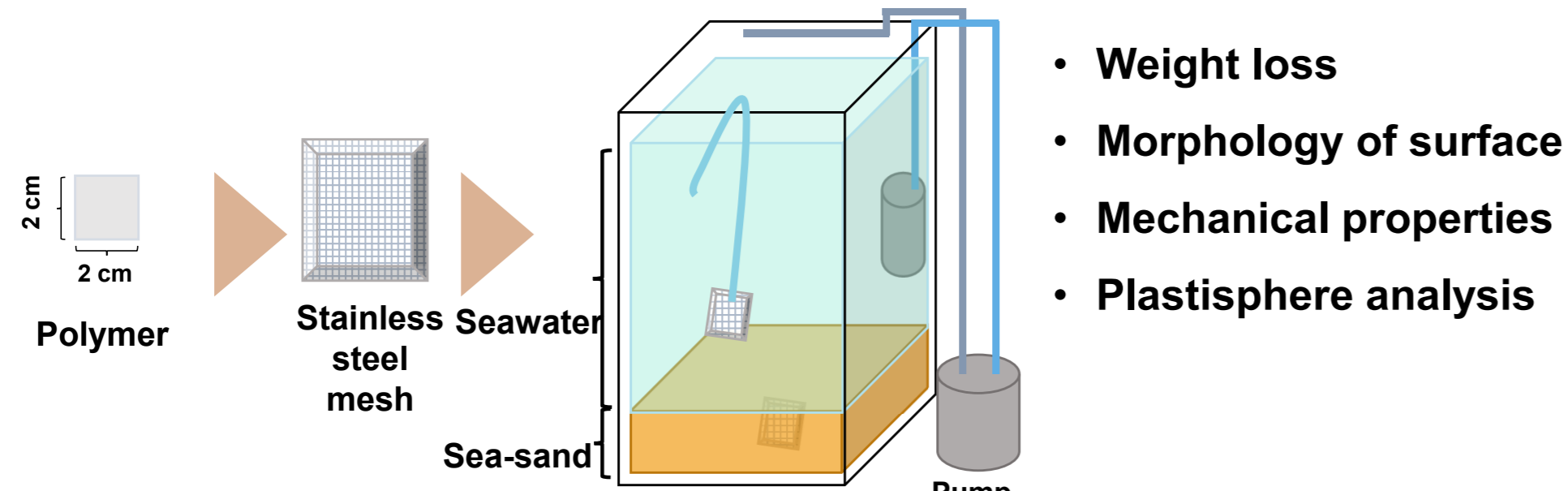


Biodegradation properties of bacteria A-F (red: high amount)

bacteria	zone	monomer	dimer	trimer	tetramer
A	+	+	+	+	+
B	+	+	+	+	+
C	+	+	+	+	+
D	+	+	+	+	+
E	+	+	+	+	+
F	+	+	+	+	+

clear zone development: -, CO₂ production: +

• Degradation property of each degrader has been analyzed by clear zone development, CO₂ production and accumulated degradation products.



Tank experiment

- Weight loss
- Morphology of surface
- Mechanical properties
- Plastisphere analysis